

HISTOLOGICAL PREPARATION

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PAPER VIII)

Collection of the Sample:

- a) Living organism are sacrificed to take the organs – heart, liver, kidney, testes, ovary, intestine, stomach etc.

Fixation of the Sample:

- a) Fixation process must be started quickly as possible as the removal of selected organs.
- b) Take 3-5 mm thick tissue/organs for quick fixation in fixatives.
- c) Plastic sample jars for fixation are labelled with sample name, collection date etc. Fill the vials with 2/3 of fixatives.
- d) Organs and fixative volume must be 1:20.
- e) Place the organs into the separate vials with fixative.
- f) Store the vial to fix the tissue. Fixation time will depend on the type of fixatives used. Fixation time – Formalin: 10-12 hrs. Alcohol formalin 7-10 hrs. and Bouin's: 4-6 hrs. (At normal room temperature).

Dehydration of the Sample:

After proper fixation, the tissue must be dehydrated to remove the water from tissue block

by following ways.

- | | |
|-----------------|--------|
| - Alcohol (50%) | - 1 hr |
| - Alcohol (70%) | - 1 hr |
| - Alcohol (70%) | - 1 hr |
| - Alcohol (85%) | - 1 hr |
| - Alcohol (95%) | - 1 hr |

- Alcohol (100%) - 1 hr

Cleaning:

After dehydration, the tissues are to be cleared with clearing agent to remove alcohol prior to paraffin embedding. The clearing agent should be miscible both in alcohol and paraffin. The commonly used cleaning agents are:

- Xylene = (3-4 mm thick tissue can be cleared in 30-60 minutes for two changes each)
- Chloroform = (Clearing time – Overnight, but less harmful than xylene for the tissue)
- Cedar wood oil (Histological GR) = (It is good for cleaning of delicate tissue).

Impregnation with paraffin:

- a) Paraffin are melted in hot air oven at 50-60 degree centigrade depending upon the melting point.
- b) Avoid to exceed the temperature 5 degree centigrade above the melting point as it can cause the tissue hardening or shrinkage. If the wax is over heated it can be crystallised and become useless.
- c) The tissue is transferred to molten wax. The amount of wax should be 25-30 times more than tissue volume.
- d) Impregnation must be with 2-3 changes in melting wax to complete removal of clearing agent.

Embedding:

It is the orientation of tissue in melted paraffin which after solidification can be used for tissue sectioning in microtome.

- a) Molten paraffin wax which is heated at temperature just 2-3 degree above the melting point is to poured into the mould to an adequate depth so as to cover the thickest tissue block.

- d) On exposing a suitable area of tissue, the section thickness is set to the appropriate level for routine purposes to 4-6 microns.
- e) Apply ice to the surface of the block for a few seconds and wipe the surface of block free of water. This step is optional but makes sections cut easily.
- f) The whole surface of the block will move parallel to the edge of the knife in order to ensure a straight ribbon of sections.
- g) The microtome is now moved in an easy rhythm with right hand operating the microtome and left hand holding the sections away from the knife.
- h) The ribbon is formed due to the slight heat generated during cutting, which causes the edges of the sections to adhere. If difficulty is experienced in forming the ribbon it is sometimes overcome by rubbing one of the edges of the block with finger.
- i) During cutting the paraffin wax embedded sections become slightly compressed and creased. Before being attached to slides the creases must be removed and the section flattened. This is achieved by floating them on warm water at 45-degree C, with little trace of detergent. It is helpful to flatten the ribbon and can remove any shrinkage.
- j) The action in floating out must be smooth with the trailing end of ribbon making contact with water first to obtain flat sections with correct orientation, floating out with the shiny surface towards the water is essential. When the ribbon has come to rest on water the remaining wrinkles and folds are removed by teasing apart by using forceps or seeker.
- k) Picking up sections – The ribbon of sections floating on water is split into individual or groups of sections by use of forceps or seekers. Picking up a section on slide is achieved by immersing the slide lightly smeared with adhesive vertically to three fourths of its length bringing the section in contact with the slide. On lifting the slide vertically from the water, the section will flatten on to the slide. The sections are then blotted lightly with moistened blotting paper to remove excess water and to increase contact between section and slide. For delicate tissues or when several ribbons of sections are placed on the slide, omit the blotting instead keep the slide in upright position for several minutes to drain.

- l) Sections are collected from the very first cut that includes any tissue. Ribbons of ten - 1-10, 11-20, 21-30 so on are picked up and mounted on the slides.
- m) Adhesive are placed on slide and rub over it before placing the section cutting.
- n) Drying of section: Sections are then kept in incubator with a temperature 5-6°C above the melting point of wax i.e. at 60°C for 20-60 minutes. It is better to overheat than underheat. If the sections are not well dried they may come off during staining. The sections should not be allowed to dry without a good contact with the slide, such sections will come off during staining.

Staining:

The process of staining in copulin jar with Haematoxylin and Eosin are as follows:

a. Cleaning and Rehydration –

- Alcohol (85%) - 30 sec.
- Alcohol (70%) - 30 sec.
- Tap water - 30 sec

b. Staining –

- Haematoxylin - 2 minutes
- Tap water - 30 sec.
- Tap water - 30 sec.
- Tap water - 30 sec.
- Alcohol (70%) - 1 minute
- Alcohol (95%) - 1 minute
- Eosin Y - 1 minute
- Alcohol (95%) - 2 minute
- Alcohol (95%) - 1 minute
- Alcohol (100%) - 2 minute
- Alcohol (100%) - 2 minute
- Alcohol (100%) - 2 minute
- XyloXylo - 1 minute minutes.
- Xylo - 1 minute
- Xylo - 1 minute
- Alcohol (100%) - 30 sec.

- Xylol - 1 minute
- Xylol - 1 minute

Mounting:

- a) The final step in this procedure is to permanently mount the sections under a coverslip.
- b) This is accomplished by covering the section in a medium that will harden and produce a clear binder between the slide and cover slip. The ideal mounting medium should not distort the stain colour, or yellow and become brittle with age. DPX or Canada balsam can be used as good mounting adhesive.
- c) Place 2-3 drops of resin over the slide carefully to avoid trapping any bubble inside it. To avoid entrapping air bubbles, lower the cover slip slowly from one side of the droplet.
- d) Place the slide on the slide warmer and carefully place a lead weight on top of the cover slip. There should be enough mounting medium to completely cover the bottom of the cover slide, and budge slightly around the edges.
- e) Leave slides on the warmer for at least 24 hours; excess medium can then be cut from edges of cover slip with a razor blade.

Fixatives:

Formalin (v/v) (10%)

- Formaldehyde= 10 ml
- Acid sodium phosphate (monohydrate) = 0.4 gm
- Anhydrous disodium phosphate= 0.65 gm
- Water to make the volume= 100 ml

Alcohol formalin (v/v) (10%)

- Formalin = 10 ml
- Alcohol (70%-95%) = 90 ml

Bouin's Fluid

- Saturated aqueous picric acid = 75 ml
- Formalin = 25 ml
- Acetic acid = 05 ml

Section adhesive Mayer's Albumin –

- Fresh egg white = 50 ml
- Glycerol = 10 ml
- Sodium salicylate = 01 ml

Stain:

Haematoxylin:

- Haematoxylin powder (Harris haematoxylin) = 4 g.
 - Potash alum = 100g.
 - Mercuric oxide = 2 g.
 - Ethanol (100%) = 60 ml.
 - Distilled water = 800 ml
- a) Boil 800 ml. distilled water with potash alum (100 g.)
 - b) Add 4 g. Haematoxylin with 60 ml ethanol to dissolve in it.
 - c) When potash dissolve in water, add ethanol haematoxylin solution in it.
 - d) After adding Haematoxylin at low heat add mercuric oxide and heat the mixture again to dissolve Mercuric oxide completely. Now the colour of the solution become pinkish blue.
 - e) In working solution of Haematoxylin (100 ml.) 4 ml of glacial acetic acid can be added for good nuclear staining.

Eosin:

- Eosin = 2g.

- Distilled water = 160 ml.
 - Alcohol (95%) = 640 ml.
 - Glacial acetic acid. = 0.8 ml.
- a) Add eosin to distilled water.
 - b) Add 95% alcohol in eosin + distilled water.
 - c) To final mixture add 0.8 ml glacial acetic acid.